



# Biphasic effect of hydrogen peroxide on field potentials in rat hippocampal slices

Hiroshi Katsuki \*, Chiaki Nakanishi, Hiroshi Saito, Norio Matsuki

Laboratory of Chemical Pharmacology, Graduate School of Pharmaceutical Sciences, The University of Tokyo, Bunkyo-ku, Tokyo 113, Japan
Received 24 April 1997; revised 19 August 1997; accepted 2 September 1997

#### Abstract

In the CA1 region of rat hippocampal slices,  $H_2O_2$  (0.294–2.94 mM) caused initial augmentation, and subsequent long-lasting depression, of population spikes and excitatory postsynaptic potentials. The effect of  $H_2O_2$  may not be mediated by its degradation product, hydroxyl radicals, because an iron chelator deferoxamine did not block the effect. A catalase inhibitor 3-amino-1,2,4-triazole only modestly attenuated the initial augmentation, suggesting that the effect of  $H_2O_2$  is not attributable to catalase-dependent  $O_2$  generation, either. An *N*-methyl-D-aspartate receptor antagonist DL-2-amino-5-phosphonovaleric acid had no influence on the effect of  $H_2O_2$ , whereas a  $\gamma$ -aminobutyric acid type A receptor channel blocker picrotoxin attenuated long-lasting depression, indicating that  $\gamma$ -aminobutyric acid-mediated inhibition is altered during the depression phase. The initial augmentation but not subsequent depression was attenuated by a phospholipase  $A_2/C$  inhibitor 4-bromophenacyl bromide, suggesting the involvement of lipid signaling molecule(s) in the enhancement of excitatory synaptic transmission. These results suggest that  $H_2O_2$  regulates hippocampal synaptic transmission via multiple mechanisms. © 1997 Elsevier Science B.V.

Keywords: Free radical; H2O2; Hippocampus; CA1 region; Synaptic plasticity

#### 1. Introduction

Free radicals and reactive oxygen species are continuously generated in the course of normal cellular metabolism but are well controlled by intrinsic detoxifying enzyme systems and antioxidants. Under certain pathological conditions, however, this balance can be disrupted. Brain tissue is particularly vulnerable to oxidative stress because of large amounts of oxygen consumption and high contents of easily oxidized constituents such as poly-unsaturated fatty acids. Oxidative insults are implicated in many pathological processes in the brain, including acute injuries like ischemia and trauma, and more slowly progressing disorders like Parkinson's and Alzheimer's disease (Halliwell, 1992; Coyle and Puttfarcken, 1993).

On the other hand, accumulating evidence suggests that reactive oxygen species such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) play important roles in physiological cellular events as intermediates of intracellular and/or intercellular signaling (Lander, 1997). Indeed, neuronal functions assessed by

electrophysiological measurements are shown to be directly affected by reactive oxygen species (Pellmar, 1987; Pellmar et al., 1989; Muller et al., 1993; Seutin et al., 1995). Furthermore, several species such as nitric oxide and carbon monoxide are implicated in the underlying mechanisms of long-lasting changes in synaptic transmission, candidate processes of information storage during learning (Zhuo et al., 1993). In the present study we describe novel, complex actions of  $H_2O_2$  on evoked excitatory field potentials in the CA1 region of hippocampal slices.

### 2. Materials and methods

Hippocampal slices were prepared from young adult (4–7 weeks old) male Wistar rats. After decapitation, the brain was rapidly removed, hippocampi were dissected and placed in gassed (95%  $\rm O_2/5\%~CO_2$ ) standard extracellular solution containing 124 mM NaCl, 5 mM KCl, 2.4 mM CaCl<sub>2</sub>, 1.3 mM MgSO<sub>4</sub>, 1.24 mM KH<sub>2</sub>PO<sub>4</sub>, 26 mM NaHCO<sub>3</sub> and 10 mM D-glucose. Transverse slices (400–450  $\mu$ m thick) were cut with a microslicer DTK-2000

<sup>\*</sup> Corresponding author. Tel.: (81-3) 3812-2111, ext. 4785; Fax: (81-3) 3815-4603

(Dosaka EM, Kyoto, Japan). Slices were then maintained in an incubation chamber for at least 1 h at 37°C in the standard solution. At the time of an experiment, individual slices were transferred to a submersion recording chamber where they were constantly perfused with the standard solution (2.5 ml/min) at 37°C.

Extracellular recordings were obtained from the pyramidal cell layer of the CA1 region using 5 to 10 M $\Omega$  glass electrodes filled with 0.9% NaCl. A bipolar electrode was placed on the stratum radiatum to stimulate Schaffer collateral/commissural pathway. Stimuli of 50 µs in duration were applied every 30 s. The stimulus intensity was set to evoke 50% of the maximal amplitude of field population spikes. In several experiments, field excitatory postsynaptic potentials (EPSPs) were recorded with a glass electrode placed on the apical dendritic layer of the CA1 region. Recorded responses were amplified, temporarily stored on a storage unit EA-602J (Nihon Kohden, Tokyo, Japan) and written out on a chart recorder. The amplitude of population spikes or EPSPs was measured from the chart paper using a digitizing tablet. Data are presented as mean  $\pm$  S.E.M.

DL-2-Amino-5-phosphonovaleric acid (APV), 3-amino-1,2,4-triazole and deferoxamine were obtained from Sigma (St. Louis, MO, USA). All other reagents and chemicals were obtained from Wako (Osaka, Japan).

#### 3. Results

Fig. 1 shows the effects of  $\rm H_2O_2$  on population spikes recorded from the CA1 pyramidal cell layer, and EPSPs recorded from the CA1 apical dendritic layer. Bath application of  $\rm H_2O_2$  (1.47 mM; 0.005%) for 20 min induced biphasic changes in the amplitude of population spikes. The field responses were initially augmented, which continued in a steady-state level during the presence of  $\rm H_2O_2$  in the perfusing solution. After washout of  $\rm H_2O_2$ , the responses were promptly depressed and gained a new steady-state level below the baseline (Fig. 1A and B). Similar results were obtained when the amplitude of EPSP was measured in the same experimental conditions (Fig. 1A and C).

These biphasic effects were evident when  $\rm H_2O_2$  was applied at concentrations of 0.294 mM or higher. The initial augmenting effect was most prominent at 1.47 mM  $\rm H_2O_2$ ; higher or lower concentrations of  $\rm H_2O_2$  resulted in smaller degrees of augmentation (Fig. 2A). On the other hand, the magnitude of subsequent depression was similar at a range of 0.294–2.94 mM. Depression caused by 0.294 mM  $\rm H_2O_2$  was partially reversible after 30 min of washout, although higher concentrations of  $\rm H_2O_2$  induced stable, long-lasting depression of population spikes (Fig. 2B).

We tested the influences of duration of  $H_2O_2$  application. When  $H_2O_2$  (1.47 mM) was perfused for 10 min, subsequent depression of population spikes was transient,

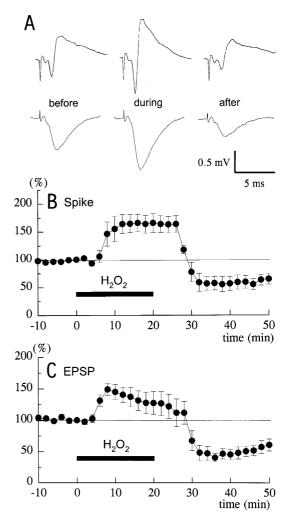


Fig. 1. (A) Representative traces of population spikes (upper) and EPSPs (lower) before, during and 30 min after application of 1.47 mM  $\rm H_2O_2$  for 20 min. (B) and (C) Time-courses of the changes in the amplitudes of population spikes (B, n=5) and EPSPs (C, n=8) induced by bath application of 1.47 mM  $\rm H_2O_2$ .  $\rm H_2O_2$  was applied during the time indicated by bars.

and the amplitude showed recovery within 30 min after washout. More prolonged (over 20 min) perfusion resulted in permanent depression of the responses (Fig. 3A). When  $\rm H_2O_2$  was applied for 40 min or longer, depression of the population spike was already evident during the prolonged perfusion of  $\rm H_2O_2$  (Fig. 3B), suggesting that the depression phase is not dependent on the removal of  $\rm H_2O_2$ .

In the next set of experiments, we examined whether the effects of  $\rm H_2O_2$  are mediated by its degradation products. Hydroxyl radicals can be generated through the Fenton reaction;  $\rm H_2O_2$  reacts with  $\rm Fe^{2+}$  to produce this extremely reactive species responsible for covalent modification and damages to macromolecules (Coyle and Puttfarcken, 1993). If hydroxyl radicals are involved, chelation of endogenous  $\rm Fe^{2+}$  by a potent iron chelator deferoxamine should alter the effect of  $\rm H_2O_2$ . However, pretreatment of slices with 20  $\mu \rm M$  deferoxamine failed to block the effect

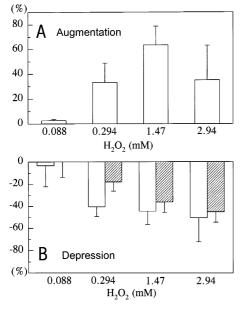
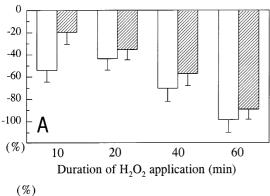


Fig. 2. Concentration dependence of the effects of  $\rm H_2O_2$  on the population spike amplitude.  $\rm H_2O_2$  was applied for 20 min at indicated concentrations. Values shown are percent increases observed at the end of  $\rm H_2O_2$  application (A) and percent decreases observed at 10 min (open columns) and 30 min (hatched columns) after washout (B). n=4-5 for each condition.



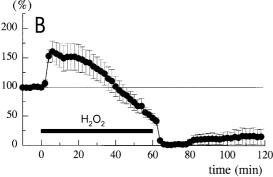


Fig. 3. Prolonged application of  $H_2O_2$  results in large magnitudes of depression. (A)  $H_2O_2$  (1.47 mM) was applied for indicated durations. Values are percent decreases in population spike amplitude observed at 10 min (open columns) and 30 min (hatched columns) after washout of  $H_2O_2$ . n=4-5 for each condition. (B) Time-course of the changes in population spike amplitude induced by 60 min application of 1.47 mM  $H_2O_2$ , n=5.

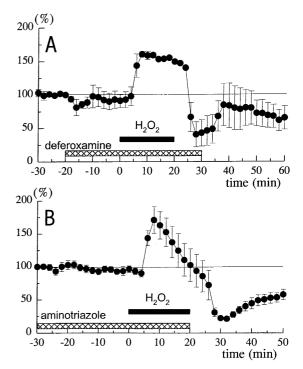


Fig. 4. Effects of deferoxamine (A, n = 4) and 3-amino-1,2,4-triazole (B, n = 7) on H<sub>2</sub>O<sub>2</sub> (1.47 mM)-induced changes in population spike amplitude. Deferoxamine (20  $\mu$ M) and 3-amino-1,2,4-triazole (10 mM) were applied during the time indicated by a hatched bar.

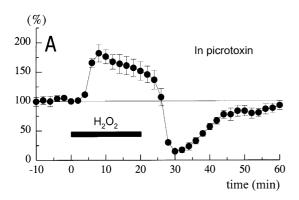
of  $H_2O_2$ . Both augmenting and depressing actions of  $H_2O_2$  on population spikes were similarly observed in the presence of deferoxamine (Fig. 4A).

Another possibility is that  $H_2O_2$  may support synaptic transmission by serving as a source of molecular oxygen O<sub>2</sub> (Llinas and Sugimori, 1980). This is due to the property of H<sub>2</sub>O<sub>2</sub>, depending on catalase activity intrinsic to brain tissues (Walton and Fulton, 1983), to be degraded into O<sub>2</sub> and H<sub>2</sub>O. Therefore, we used a catalase inhibitor 3-amino-1,2,4-triazole to investigate whether the actions of H<sub>2</sub>O<sub>2</sub> on population spikes were mediated by catalase-dependent  $O_2$  generation from  $H_2O_2$ . As shown in Fig. 4B, the only difference observed in slices treated with 3amino-1,2,4-triazole was that augmentation during H<sub>2</sub>O<sub>2</sub> application was not persistent and the amplitude of population spikes gradually declined to the level at or below the baseline. However, the magnitude of initial augmentation at the beginning of  $H_2O_2$  application and that of enduring depression after the washout of H<sub>2</sub>O<sub>2</sub> were not significantly different from those observed in control slices (Fig. 1B). These results indicate that the effects of  $H_2O_2$  are not attributable to catalase-dependent O<sub>2</sub> generation, although endogenous catalase does modulate the actions of H<sub>2</sub>O<sub>2</sub>.

The modulation of evoked field potentials could result from alteration of transmission efficacy either at excitatory synapses or at inhibitory synapses. To clarify these alternatives,  $\rm H_2O_2$  application was performed in the presence of picrotoxin (50  $\mu$ M), a blocker of GABA<sub>A</sub> receptor chan-

nels. As shown in Fig. 5A, picrotoxin had no effect on the augmentation of population spikes during application of  $H_2O_2$ . In contrast, depression observed after washout of  $H_2O_2$  was affected by picrotoxin. Although the transient component of depression shortly after  $H_2O_2$  washout was accelerated, the sustained component long after washout was attenuated. The amplitude of population spikes gradually returned to the level near the baseline. Thus,  $H_2O_2$ -induced augmentation may be the consequences of the enhancement of excitatory synaptic transmission, whereas long-lasting depression by  $H_2O_2$  may involve alteration of transmission efficacy at inhibitory synapses.

The augmentation by  $\rm H_2O_2$  seems to be different from long-term potentiation of synaptic transmission, a well-known form of synaptic plasticity at hippocampal excitatory synapses. First, increases in field responses were observed only during application of  $\rm H_2O_2$  and disappeared promptly after washout, which is not consistent with the characteristic persistence of electrically-induced long-term potentiation. Second, although the activation of NMDA receptors is critical for the induction of long-term potentiation by electrical stimulation in CA1 synapses (Bliss and Collingridge, 1993), a selective NMDA receptor antagonist APV (50  $\mu$ M) showed no effect on the magnitude of augmentation induced by  $\rm H_2O_2$  (Fig. 5B). In addition, the



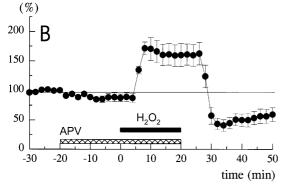


Fig. 5. Effects of picrotoxin (A, n=5) and DL-2-amino-5-phosphonovaleric acid (APV) (B, n=5) on  ${\rm H_2O_2}$  (1.47 mM)-induced changes in population spike amplitude. Picrotoxin (50  $\mu$ M) was present during the entire course of experiments, whereas APV (50  $\mu$ M) was applied during the time indicated by a hatched bar.

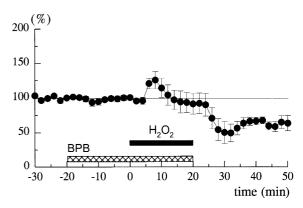


Fig. 6. Effect of 4-bromophenacyl bromide (20  $\mu$ M) on H<sub>2</sub>O<sub>2</sub> (1.47 mM)-induced changes in population spike amplitude (n=5). 4-Bromophenacyl bromide was applied during the time indicated by a hatched bar.

depression of population spikes observed after washout of  $H_2O_2$  was not altered by APV, either.

It is becoming increasingly evident that various kinds of cellular biochemical processes are influenced by reactive oxygen species. Among them is phospholipase A2, which is reported to be activated by reactive oxygen species (Goldman et al., 1992). Considering the evidence that several products of phospholipases, such as arachidonic acid and its metabolites, directly affect synaptic transmission at central synapses (Drapeau et al., 1990), we speculated that phospholipase-mediated processes may be involved in the effects of  $H_2O_2$ . Accordingly, we examined the effect of a phospholipase C/A<sub>2</sub> inhibitor 4bromophenacyl bromide on H2O2-induced changes in excitatory field responses. Slices treated with 4bromophenacyl bromide (20 µM) prior to and during the application of H<sub>2</sub>O<sub>2</sub> (1.47 mM) showed only small, transient augmentation of population spikes during the presence of H<sub>2</sub>O<sub>2</sub> (Fig. 6). In contrast, depression after washout of H<sub>2</sub>O<sub>2</sub> was not affected by treatment with 4bromophenacyl bromide.

#### 4. Discussion

The present results demonstrate that  $H_2O_2$  affects evoked field responses in the hippocampal CA1 region. The effect apparently consisted of two phases: initial augmentation and subsequent long-lasting depression.

The initial augmenting effect was not influenced by blockade of  $GABA_A$  receptor channels, indicating that it results from the enhancement of synaptic transmission at excitatory synapses. Earlier studies have shown that  $H_2O_2$  can support synaptic transmission in brain slice preparations, even in the absence of added  $O_2$  (Llinas and Sugimori, 1980; Walton and Fulton, 1983). This effect is exerted by generation of  $O_2$  from  $H_2O_2$ , and is entirely

dependent on catalase activity intrinsic to the tissues (Walton and Fulton, 1983). In this study, we observed that the inhibition of endogenous catalase by 3-amino-1,2,4-triazole prevented the persistence of augmentation during H<sub>2</sub>O<sub>2</sub> application, which suggests that endogenous catalase does play some role in controlling the modulatory effects of H<sub>2</sub>O<sub>2</sub> on synaptic transmission. However, pretreatment with 3-amino-1,2,4-triazole failed to block initial augmentation at the beginning of H<sub>2</sub>O<sub>2</sub> application. This result indicates that O<sub>2</sub> supply by catalase-dependent degradation is unlikely to be a primary mechanism of the augmenting effect of H<sub>2</sub>O<sub>2</sub>. Moreover, deferoxamine did not affect H<sub>2</sub>O<sub>2</sub>-induced augmentation, demonstrating that the hydroxyl radical is not involved in this effect. Taken together, these results suggest that H<sub>2</sub>O<sub>2</sub> itself, rather than its degradation products, is responsible for the augmentation of excitatory synaptic transmission.

On the other hand, phospholipase  $A_2/C$  inhibitor 4bromophenacyl bromide largely attenuated the augmenting effect of H<sub>2</sub>O<sub>2</sub>. In several preparations, reactive oxygen species including H<sub>2</sub>O<sub>2</sub> have been demonstrated to activate phospholipase A<sub>2</sub> (Goldman et al., 1992; Rao et al., 1993). Lipid signaling molecules such as arachidonic acid and platelet-activating factor are generated by phospholipase A<sub>2</sub> activity, and can regulate the efficacy of hippocampal synaptic transmission (Drapeau et al., 1990; Clark et al., 1993). Therefore, it is likely that these lipid mediators are involved in the augmentation of synaptic responses induced by H<sub>2</sub>O<sub>2</sub>. In this context, increase in spontaneous miniature excitatory postsynaptic currents induced by a brief period of anoxia is blocked by 4bromophenacyl bromide (Katchman and Hershkowitz, 1994), suggesting that phospholipase products, possibly arachidonic acid and/or its metabolites, can augment excitatory synaptic transmission.

Depression observed after washout of H<sub>2</sub>O<sub>2</sub> was longlasting, when sufficient concentrations of H<sub>2</sub>O<sub>2</sub> and durations of application were employed. This long-lasting depression was not attenuated either by an iron chelator or by a catalase inhibitor, leaving the possibility that it is not due to non-specific deterioration of tissue viability caused by oxidative injury. It is interesting to note that H<sub>2</sub>O<sub>2</sub> induces a long-lasting inhibition of Ca<sup>2+</sup>-dependent glutamate release from cerebrocortical synaptosomes even in the presence of deferoxamine, therefore, apparently without damaging the synaptosomes (Zoccarato et al., 1995). In addition, picrotoxin blocked the persistence of depression, indicating that alterations in GABA-mediated inhibitory control are at least in part involved in the long-lasting depression of synaptic responses observed after H<sub>2</sub>O<sub>2</sub> treatment. The exact mechanisms still remain unclear and require further clarification: although arachidonic acid or its metabolites are reported to participate in long-lasting depression of synaptic transmission under certain conditions (Bolshakov and Siegelbaum, 1995; Normandin et al., 1996), this may not be the case with the present observations because 4-bromophenacyl bromide had no effect on  $\rm H_2O_2$ -induced depression.

Our present results are not consistent with those reported by Pellmar and colleagues, who examined the effects of H<sub>2</sub>O<sub>2</sub> on field potentials in the CA1 region of guinea pig hippocampal slices (Pellmar, 1987; Pellmar et al., 1989). First, they have not described an augmenting action of  $H_2O_2$ . Second, although they found that  $H_2O_2$ decreased synaptic responses, in their studies deferoxamine blocked the effect of  $H_2O_2$ , leading them to the conclusion that hydroxyl radicals are responsible for this effect. Reasons for these discrepancies are not clear: besides the difference in species of animals, several subtle differences in experimental conditions such as temperature (30°C vs. 37°C in the present study) may be taken into consideration. For example, the involvement of nitric oxide in the induction of hippocampal synaptic plasticity is reported to be strictly dependent on temperature (Williams et al., 1993).

In conclusion, we have demonstrated that synaptic transmission at hippocampal CA1 synapses is significantly modulated by  $H_2O_2$ . This phenomenon may be relevant to pathological processes where reactive oxygen species are massively produced, and physiological processes such as information storage where enduring changes in transmission efficacy are involved.

## References

Bliss, T.V., Collingridge, G.L., 1993. A synaptic model of memory: Long-term potentiation in the hippocampus. Nature 361, 31–39.

Bolshakov, V.Y., Siegelbaum, S.A., 1995. Hippocampal long-term depression: Arachidonic acid as a potential retrograde messenger. Neuropharmacology 34, 1581–1587.

Clark, G.D., Happel, L.T., Zorumski, C.F., Bazan, N.G., 1993. Enhancement of hippocampal excitatory synaptic transmission by platelet-activating factor. Neuron 9, 1211–1216.

Coyle, J.T., Puttfarcken, P., 1993. Oxidative stress, glutamate, and neurodegenerative disorders. Science 262, 689–695.

Drapeau, C., Pellerin, L., Wolfe, L.S., Avoli, M., 1990. Long-term changes of synaptic transmission induced by arachidonic acid in the CA1 subfield of the rat hippocampus. Neurosci. Lett. 115, 286–292.

Goldman, R., Ferber, E., Zort, U., 1992. Reactive oxygen species are involved in the activation of cellular phospholipase A<sub>2</sub>. FEBS Lett. 309, 190–192.

Halliwell, B., 1992. Reactive oxygen species and the central nervous system. J. Neurochem. 59, 1609–1623.

Katchman, A.N., Hershkowitz, N., 1994. Arachidonic acid participates in the anoxia-induced increase in mEPSC frequency in CA1 neurons of the rat hippocampus. Neurosci. Lett. 168, 217–220.

Lander, H.M., 1997. An essential role for free radicals and derived species in signal transduction. FASEB J. 11, 118–124.

Llinas, R., Sugimori, M., 1980. Electrophysiological properties of in vitro Purkinje cell somata in mammalian cerebellar slices. J. Physiol. (Lond.) 305, 171–195.

Muller, M., Fontana, A., Zbinden, G., Gahwiler, B.H., 1993. Effects of interferons and hydrogen peroxide on CA3 pyramidal cells in rat hippocampal slice cultures. Brain Res. 619, 157–162.

Normandin, M., Gagne, J., Bernard, J., Elie, R., Miceli, D., Baudry, M., Massicotte, G., 1996. Involvement of the 12-lipoxygenase pathway of arachidonic acid metabolism in homosynaptic long-term depression of the rat hippocampus. Brain Res. 730, 40–46.

- Pellmar, T.C., 1987. Peroxide alters neuronal excitability in the CA1 region of guinea-pig hippocampus in vitro. Neuroscience 23, 447–456.
- Pellmar, T.C., Neel, K.L., Lee, K.H., 1989. Free radicals mediate peroxidative damage in guinea pig hippocampus in vitro. J. Neurosci. Res. 24, 437–444.
- Rao, G.N., Lassegue, B., Griendling, K.K., Alexander, R.W., 1993. Hydrogen peroxide stimulates transcription of c-jun in vascular smooth muscle cells: Role of arachidonic acid. Oncogene 8, 2759–2764.
- Seutin, V., Scuvee-Moreau, J., Massotte, L., Dresse, A., 1995. Hydrogen peroxide hyperpolarizes rat CA1 pyramidal neurons by inducing an increase in potassium conductance. Brain Res. 683, 275–278.
- Walton, K., Fulton, B., 1983. Hydrogen peroxide as a source of molecular oxygen for in vitro mammalian CNS preparations. Brain Res. 278, 387–393.

- Williams, J.H., Li, Y.-G., Nayak, A., Errington, M.L., Murphy, K.P., Bliss, T.V., 1993. The suppression on long-term potentiation in rat hippocampus by inhibitors of nitric oxide synthase is temperature and age dependent. Neuron 11, 877–884.
- Zhuo, M., Small, S.A., Kandel, E.R., Hawkins, R.D., 1993. Nitric oxide and carbon monoxide produce activity-dependent long-term synaptic enhancement in hippocampus. Science 260, 1946–1950.
- Zoccarato, F., Valente, M., Alexandre, A., 1995. Hydrogen peroxide induces a long-lasting inhibition of the Ca<sup>2+</sup>-dependent glutamate release in cerebrocortical synaptosomes without interfering with cytosolic Ca<sup>2+</sup>. J. Neurochem. 64, 2552–2558.